



Development of a physiologically-based pharmacokinetic model for cyclosporine in Asian children with renal impairment

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This study aimed to assess the pharmacokinetics of cyclosporine A (CsA) in Asian children with renal impairment (RI) by developing a physiologically-based pharmacokinetic (PBPK) model with Simcyp Simulator. The PBPK model of Asian children with RI was developed by modifying the physiological parameters of the built-in population libraries in Simcyp Simulator. The ratio of healthy and RI populations was obtained for each parameter showing a difference between the populations. Each ratio was multiplied by the corresponding parameter in healthy Asian children. The model verification was performed with published data of Korean children with kidney disease given multiple CsA administrations. Simulations were performed with different combinations of ethnicity, age, and renal function to identify the net impact of each factor. The simulated results suggested that the effect of RI was higher in children than adults for both Caucasian and Asian. In conclusion, the constructed model adequately characterized CsA pharmacokinetics in Korean children with RI. Simulations with populations categorized by ethnicity, age, and renal function enabled to assess the net impact of each factor on specific populations.

Introduction

Cyclosporine A (CsA), a calcineurin inhibitor, is a commonly prescribed immunosuppressant for glomerular disease in children.[1,2] Even though its primary target organ is kidney, CsA has nephrotoxicity, which requires caution when using the drug. According to the 'Kidney Disease: Improving Global Outcomes (KDIGO) Clinical Practice Guideline for Glomerulonephritis', CsA is not suggested for patients with an estimated glomerular filtration rate (GFR) below 30 mL/min/1.73 m², which is classified as severe renal impairment.[3] For patients with moderate

renal impairment (GFR ranging 30–60 mL/min/1.73 m²), CsA is used following a rigorous evaluation of the risks and benefits. Due to its dose-related nephrotoxicity, maintaining the therapeutic concentration range is essential.[4] However, its narrow therapeutic range and high inter-individual and intra-individual variability in pharmacokinetics (PK) make it challenging to maintain the therapeutic concentration range which, in turn, may lead to poor clinical outcomes.[4,5]

In addition, there are general sources of variability in PK of CsA, which adds complications to providing appropriate therapy. CsA is primarily metabolized by cytochrome P-450 3A (CYP3A) and is a substrate of multidrug resistance efflux transporter, P-glycoprotein (P-gp). Previous studies have shown that polymorphisms in genes encoding CYP3A and P-gp may affect PK of drugs.[6] Ethnicity, linked to genetic polymorphisms of CYP3A and P-gp, has been reported to influence the PK profile of CsA.[7,8] The concomitant disease could affect the PK

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of the drug. Although only 0.1% of CsA is eliminated via the kidney,[9] not only hepatic impairment but also renal impairment affects the metabolism of the liver and eventually result in PK alterations. Accumulated uremic toxins in renally impaired patients are associated with alteration of the transporters and metabolic enzymes by genetic modification.[10,11] Age is another known source of PK variability of CsA.[12] Despite its high utility in children, there are limited sources describing the pediatric PK of CsA.[13-15] In accordance with labeling information, the pediatric dosage for transplantation or renal disease is adjusted by one's body weight.[16] However, CsA dose based on body weight alone does not efficiently achieve concentration targets in children.[17] Inter-individual variability from ethnicity, disease, and age all cause PK changes for the drug, which eventually may lead to a failed therapeutic response.[18]

Considering multiple factors are linked to PK variability, the whole-body physiologically-based pharmacokinetic (PBPK) model could be a useful tool for predicting PK of CsA. The whole-body PBPK model describes the PK of a drug by integrating known physiological information with biochemical processes and interactions.[19] Simcyp Simulator is the PBPK modeling and simulation platform that enables mechanistic and quantitative prediction of the impact of concomitant diseases, age, and even ethnicity. However, there are no details available yet on the specific ethnicity of children with renal impairment in the software, which limits PK prediction within the relevant population. This study primarily aimed to predict PK of CsA in Asian (Korean and Japanese) children with renal impairment by developing a PBPK model for the corresponding population. In addition, the net effect of ethnicity, age, and renal function on PK of CsA was assessed with the developed PBPK models.

Methods

Initial PBPK model development for specific populations

To assess the PK of CsA of Asian children with renal impairment, the PBPK models of Asian children with moderate and severe renal impairment were developed. The severity of renal impairment was based on GFR. A GFR ranging from 30 to 60 mL/min/1.73 m² was considered 'moderate renal impairment', and a GFR less than 30 mL/min/1.73 m² was considered 'severe renal impairment'.

The whole-body PBPK model was developed using the Simcyp Simulator (Version 14 Release 1, Simcyp Ltd, a Certara Company, Sheffield, UK). Drug-specific parameters – physicochemical, blood binding, absorption, distribution, elimination, interaction, and transport – were taken from the compound file of the Simcyp Simulator. Table 1 presents the compound-related parameters integrated into this study. The physiological parameters were based on predetermined parameters of the Simcyp built-in population libraries. The built-in population libraries used in this study were as follows: 'Sim-Healthy Volunteers' for healthy Caucasians, 'Sim-RenalGFR30-60' and 'Sim-RenalGFR

Table 1. Physicochemical properties and pharmacokinetic parameters of cyclosporine A used for the development of the PBPK model

| Parameter | Input value |
|--|-----------------|
| Physicochemical properties | |
| Molecular weight (g/mol) | 1202.000 |
| log <i>P</i> | 2.960 |
| Compound type | Small molecule |
| Blood-to-plasma partition ratio | 1.620 |
| Hematocrit | 45.000 |
| Fraction unbound in plasma | 0.036 |
| Absorption | |
| Absorption model | First order |
| Absorption rate constant (h ⁻¹) | 1.659 |
| Lag time (h)* | 0.576 |
| Caco-2 cell permeation (10 ⁻⁶ cm/s) | 17.000 |
| Distribution | |
| Distribution model | Full PBPK model |
| V _{ss} (L/kg) | Predicted** |
| K _p scalar | 1.000 |
| Elimination | |
| Clearance type | Enzyme kinetics |
| Renal clearance (L/h) | 0.029 |
| Active uptake into hepatocyte | 1.534 |
| Hepatic clearance (μL/min/10 ⁶) | 0.690 |

All parameters except lag time are predetermined values of the Simcyp Simulator.

*Built-in substrate model of CsA (1 h) was optimized based on the literature.

**Rodgers and Rowland prediction method was used.[27]

Abbreviations: *P*, octanol-water partition coefficient; PBPK, physiologically-based pharmacokinetic; V_{ss}, volume of distribution at steady-state; K_p, partition coefficient.

less than 30', which were based on North European Caucasian data, for Caucasians with moderate and severe renal impairment, respectively, and 'Sim-Japanese' for healthy Asians. For each population, a pediatric module is provided in the software. Changes in physiological parameters with age, such as ontogeny of drug-metabolizing enzymes, are applied to the simulations with a pediatric module.

To determine the physiological parameters that need to be modified, parameter values of healthy Caucasian adults were compared with those of Caucasian adults with renal impairment. The ratio of the healthy Caucasian adult population and the renal impairment population was calculated for each parameter that was different between the populations (renal impairment/healthy). Each ratio was then multiplied by the corresponding parameter for healthy Asian children.

Model verification

The constructed model was verified by observed clinical data

from previously published literature.[20] Clinical data were comprised of PK data from 34 Korean children of age ranging from 2.3 to 17 (mean±standard deviation: 8.7±4.0 years) with nephrotic syndrome or glomerular diseases (male-female ratio of 2.2:1). Baseline mean creatinine clearance was 97.5±22.3 mL/min/1.73m². Either capsules or the syrup formulation of CsA microemulsion (Cipol-N, Chong Keun Dang, Seoul, Republic of Korea) was given at 5 mg/kg/day, twice daily. The steady-state PK parameters were calculated with plasma CsA concentrations of 3–5 days after the first dose.

The same age and dosing regimen with observed data were applied to the simulation for verification. The simulated PK profile of 0–12 h of day 5 (from 96 h to 108 h after the first dose) was observed. In addition, PK parameters, including maximum plasma concentration at steady-state ($C_{max,ss}$), area under the curve over 12 h after the last dosing ($AUC_{\tau,ss}$) and time at which $C_{max,ss}$ was observed (T_{max}), were assessed.

The predictive performance using PBPK models was determined by visual predictive check. The mean and its 5–95% confidence intervals of simulated plasma concentration-time profiles were plotted with observed data for a visual predictive check.

Comparison of PK of CsA by ethnicity, age, and renal function

To identify the net impact of ethnicity, age, and renal impairment on PK of CsA, simulations were performed with the following subpopulations:

- i. Healthy Caucasian adults and Caucasian adults with moderate and severe renal impairment.
- ii. Healthy Caucasian children and Caucasian children with moderate and severe renal impairment.
- iii. Healthy Asian adults and Asian adults with moderate and severe renal impairment.
- iv. Healthy Asian children and Asian children with moderate and severe renal impairment.

The built-in population libraries of the Simcyp Simulator were utilized for simulations of Caucasian adults (i) and children (ii). Simulations of Asian children with renal impairment (iv) were conducted with the newly developed PBPK model of this study. Using the PBPK model development method for Asian children with renal impairment, PBPK models for Asian adults with renal impairment were developed. Specifically, the ratio of the healthy Caucasian adult population and renal impairment population for parameters showing discrepancies between the populations was calculated (renal impairment/healthy). Then, each ratio was multiplied by the corresponding parameter of the healthy Asian adult population. In the built-in Simcyp population libraries, the body weight and height of the population with impaired renal function were lower than that of the healthy population. To minimize confounding factors, body weight and height (baseline and coefficient of variation) were adjusted to the same as the healthy population.

A single simulation was composed of 10 trials of 10 subjects

per trial. Age ranges for adults and children were 20–50 years and 0–15 years, respectively. All virtual individuals generated from the simulation were set as receiving 5 mg/kg/day capsule formulation, twice daily. PK parameters including $C_{max,ss}$, $AUC_{\tau,ss}$, T_{max} and overall plasma concentration-time profile were compared.

Results

Initial PBPK model development

PBPK models integrating ethnicity, age and renal impairment were developed. Several parameter values of kidney, liver and gastro-intestinal and tissue composition parts showed discrepancies between healthy and renally impaired populations in the Simcyp Simulator.

The abundance of cytochrome P450 enzymes in the liver was lower in the renal impairment populations than the healthy population. The CYP3A4 extensive metabolizers decreased 0.524-fold and 0.437-fold in moderate and severe renal impairment populations, respectively. CYP3A4-CYP3A5 correlation showed moderate differences between healthy and renal impairment populations. Parameters related to serum creatinine increased while kidney size decreased in renal impairment populations. The baseline serum creatinine increased approximately 2-fold and 4-fold, whereas baseline kidney size lowered 0.545-fold and 0.370-fold in moderate and severe renal impairment populations, respectively. Hematocrit and serum albumin values were slightly decreased in renal impairment populations. Among gastrointestinal parameters, the mean colon transit time was prolonged in the renal impairment populations. The altered physiological parameters in the renal impairment population compared to the healthy population is summarized in Table 2.

Model verification and optimization

Simulations of healthy Asian children and Asian children with renal impairment were performed with the built-in and newly developed PBPK model, respectively. Simulations with renally impaired PBPK models presented approximately 1 hour of lag time in the absorption phase. To optimize the absorption profiles, the lag time before absorption of the built-in compound file was modified. Based on the published literature, 1 h of lag time was replaced by 0.576 h.[21-23]

Simulations with final PBPK models

The final simulation results with modified lag times are shown in Figure 1. Asian children with moderate renal impairment reflected the observed data better than severe renal impairment population by means of visual predictive check. PK parameters from simulation and observed data were also compared. The T_{max} values were 1.10 h, 1.20 h, 1.25 h for healthy, moderate RI and severe RI Asian pediatric populations, respectively. The $C_{max,ss}$ and $AUC_{\tau,ss}$ tended to increase as the severity of disease increased (Fig. 2). Simulations with PBPK models of renal im-

Table 2. Summary of altered physiological parameters in the renal impairment population compared to the healthy population

| Parameters | Healthy Asian children* | Ratio from Caucasian adult population | | Renal Impairment Asian children | |
|--|-------------------------|---------------------------------------|--------------------|---------------------------------|---------|
| | | Moderate* / Healthy* | Severe* / Healthy* | Moderate | Severe |
| Liver | | | | | |
| Metabolic enzyme abundance (mean, pmol/mg) | | | | | |
| CYP1A2 EM | 44.000 | 0.546 | 0.527 | 24.031 | 23.185 |
| CYP2A6 EM | 35.000 | 0.545 | 0.470 | 19.075 | 16.450 |
| CYP2B6 EM | 3.000 | 0.547 | 0.471 | 1.641 | 1.412 |
| CYP2B6 PM | 1.000 | 0.550 | 0.467 | 0.550 | 0.467 |
| CYP2C8 EM | 14.000 | 0.546 | 0.471 | 7.642 | 6.592 |
| CYP2C9 EM | 73.000 | 0.547 | 0.473 | 39.900 | 34.500 |
| CYP2C9 PM | 29.000 | 0.548 | 0.472 | 15.900 | 13.700 |
| CYP2C18 EM | 1.000 | 0.680 | 0.570 | 0.680 | 0.570 |
| CYP2C19 EM | 1.000 | 0.543 | 0.429 | 0.543 | 0.429 |
| CYP2D6 EM | 5.000 | 0.575 | 0.450 | 2.875 | 2.250 |
| CYP2D6 UM | 9.000 | 0.575 | 0.450 | 5.175 | 4.050 |
| CYP2E1 EM | 44.000 | 0.611 | 0.423 | 26.905 | 18.610 |
| CYP2J2 EM | 2.000 | 0.725 | 0.558 | 1.450 | 1.117 |
| CYP3A4 EM** | 122.000 | 0.524 | 0.437 | 63.939 | 53.342 |
| CYP3A4-CYP3A5 Correlation (%)** | 49.980 | 0.724 | 0.555 | 36.202 | 27.755 |
| Tissue composition | | | | | |
| Blood composition** | | | | | |
| Hematocrit Mean (male) (%) | 43.000 | 0.923 | 0.772 | 39.700 | 33.200 |
| Hematocrit Mean (female) (%) | 38.000 | 0.961 | 0.824 | 36.500 | 31.300 |
| Serum Albumin (male) (g/L) | 50.340 | 0.936 | 0.856 | 47.100 | 43.080 |
| Serum Albumin (female) (g/L) | 49.380 | 0.909 | 0.765 | 44.900 | 37.800 |
| Serum Albumin C1 (female) | -0.037 | 1.000 | 1.554 | -0.037 | -0.058 |
| Kidney | | | | | |
| Serum creatinine (µmol/L)** | | | | | |
| Male Baseline | 76.500 | 1.987 | 3.922 | 152.000 | 300.000 |
| Male Baseline CV (%) | 16.100 | 0.497 | 0.497 | 8.000 | 8.000 |
| Male Age Cut-off Baseline | 81.200 | 1.761 | 3.695 | 143.000 | 300.000 |
| Male Age Cut-Off CV1 (%) | 27.400 | 0.292 | 0.292 | 8.000 | 8.000 |
| Male Age Cut-Off CV2 (%) | 21.200 | 0.377 | 0.377 | 8.000 | 8.000 |
| Female Baseline | 57.000 | 2.667 | 5.263 | 152.000 | 300.000 |
| Female Baseline CV (%) | 20.400 | 0.441 | 0.490 | 9.000 | 10.000 |
| Female Age Cut-Off 1 | 48.000 | 1.250 | 1.000 | 60.000 | 48.000 |
| Female Age Cut-Off Baseline 2 | 66.200 | 2.236 | 4.532 | 148.000 | 300.000 |
| Female Age Cut-off 1 CV 1 (%) | 26.500 | 0.340 | 0.377 | 9.000 | 10.000 |
| Female Age Cut-off 1 CV 2 (%) | 22.800 | 0.395 | 0.439 | 9.000 | 10.000 |
| Female Age Cut-Off 2 | 78.000 | 0.962 | 1.000 | 75.000 | 78.000 |
| Female Age Cut-Off Baseline 3 | 79.500 | 1.811 | 3.774 | 144.000 | 300.000 |
| Female Age Cut-Off 2 CV1 (%) | 38.300 | 0.235 | 0.261 | 9.000 | 10.000 |
| Female Age Cut-Off 2 CV2 (%) | 31.600 | 0.285 | 0.316 | 9.000 | 10.000 |
| Kidney Size | | | | | |
| Baseline | 15.400 | 0.545 | 0.370 | 8.400 | 5.700 |
| BW coefficient | 2.040 | 0.804 | 0.510 | 1.640 | 1.000 |
| BH coefficient | 51.800 | 0.633 | 0.575 | 32.800 | 29.800 |
| Gastrointestinal Tract | | | | | |
| Mean colon transit time (h) | 13.8 | 1.250 | 1.250 | 17.250 | 17.250 |

*Parameter values are from built-in model of Simcyp Simulator.

**Physiological parameters used for developing the renally impaired pediatric model in this study.

Abbreviations: CV, coefficient of variations; EM, extensive metabolizers; PM, poor metabolizers; UM, ultra-rapid metabolizers; BW, Baseline width; BH, Baseline height.

pairment population presented higher $C_{max,ss}$ and $AUC_{t,ss}$ values than the observed data.

Ethnicity, age, and renal function

Simulations were conducted for combinations of different ethnicities and age groups (Table 3, Fig. 3). Asians had lower $C_{max,ss}$ and higher $AUC_{t,ss}$ when compared with Caucasians. In the healthy and moderate renal impairment populations, $C_{max,ss}$ and $AUC_{t,ss}$ values of children were lower than those of adults. However, in severe renal impairment populations, $C_{max,ss}$ and $AUC_{t,ss}$ were higher in children than those of adults. In addition, the de-

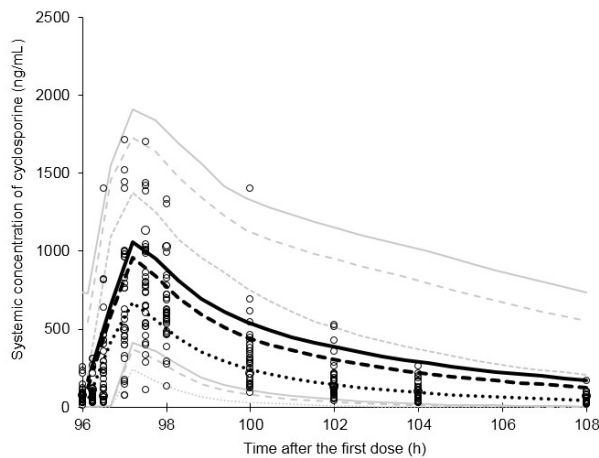


Figure 1. Overlay of observed (circles) blood concentration and simulated (lines) steady-state plasma concentration-time profile of cyclosporine A. Dotted lines indicate a healthy population. Dashed lines indicate a population with moderate renal impairment. Solid lines indicate a population with severe renal impairment. Black lines indicate the overall mean for the virtual populations. The gray lines indicate the 5–95% confidence intervals.

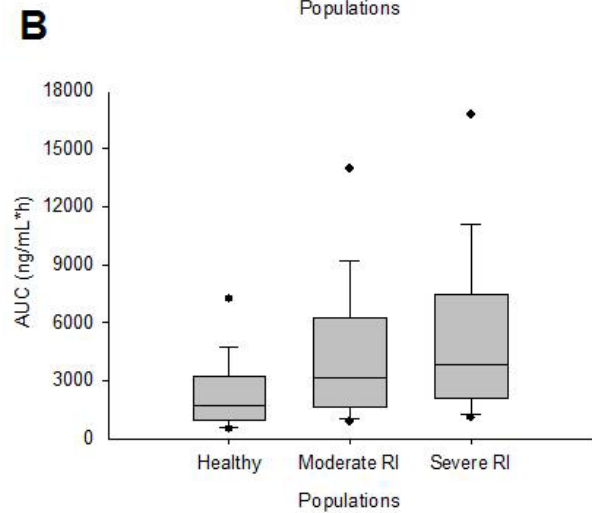
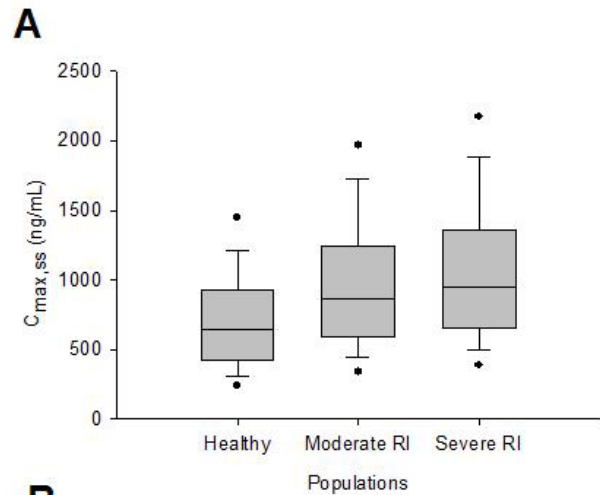


Figure 2. Comparison of simulated (A) $C_{max,ss}$ and (B) $AUC_{t,ss}$ of CsA in three populations: healthy Asian children, Asian children with moderate renal impairment and Asian children with severe renal impairment.

Table 3. PBPK model-simulated steady-state pharmacokinetic parameters of cyclosporine A for subpopulations administered 5 mg/kg/day twice daily cyclosporine A

| | Adult (20–50 years) | | Children (0–15 years) | |
|------------------|----------------------------|-----------------------------|----------------------------|-----------------------------|
| | $C_{max,ss}$ (ng/mL) | $AUC_{t,ss}$ (ng/mL·h) | $C_{max,ss}$ (ng/mL) | $AUC_{t,ss}$ (ng/mL·h) |
| Caucasian | | | | |
| Healthy | 769.13 ± 374.18 | 2752.31 ± 1950.09 | 749.25 ± 320.66 | 2294.06 ± 1463.19 |
| Moderate RI* | 1011.84 ± 494.15 (1.32) | 4842.14 ± 3536.44 (1.76) | 1016.87 ± 423.46 (1.36) | 4382.19 ± 2851.01 (1.91) |
| Severe RI* | 1009.34 ± 503.19 (1.31) | 5000.82 ± 3731.04 (1.82) | 1113.65 ± 460.79 (1.49) | 5285.23 ± 3414.46 (2.30) |
| Asian | | | | |
| Healthy | 716.02 ± 392.47 | 2776.89 ± 2398.42 | 736.85 ± 344.71 | 2333.15 ± 1868.38 |
| Moderate RI* | 945.05 ± 534.32 (1.32) | 4874.2 ± 4361.26 (1.76) | 1001.64 ± 471.57 (1.36) | 4398.79 ± 3656.58 (1.89) |
| Severe RI* | 950.49 ± 545.97 (1.33) | 5106.51 ± 4588.51 (1.84) | 1103.5 ± 520.23 (1.50) | 5324.4 ± 4376.57 (2.28) |

*Data are presented as mean ± standard deviation (ratio of RI population to healthy population RI/healthy). Abbreviations: RI, renal impairment; $C_{max,ss}$, the maximum plasma concentration at steady-state; $AUC_{t,ss}$, area under the curve over 12 h after the last dosing.

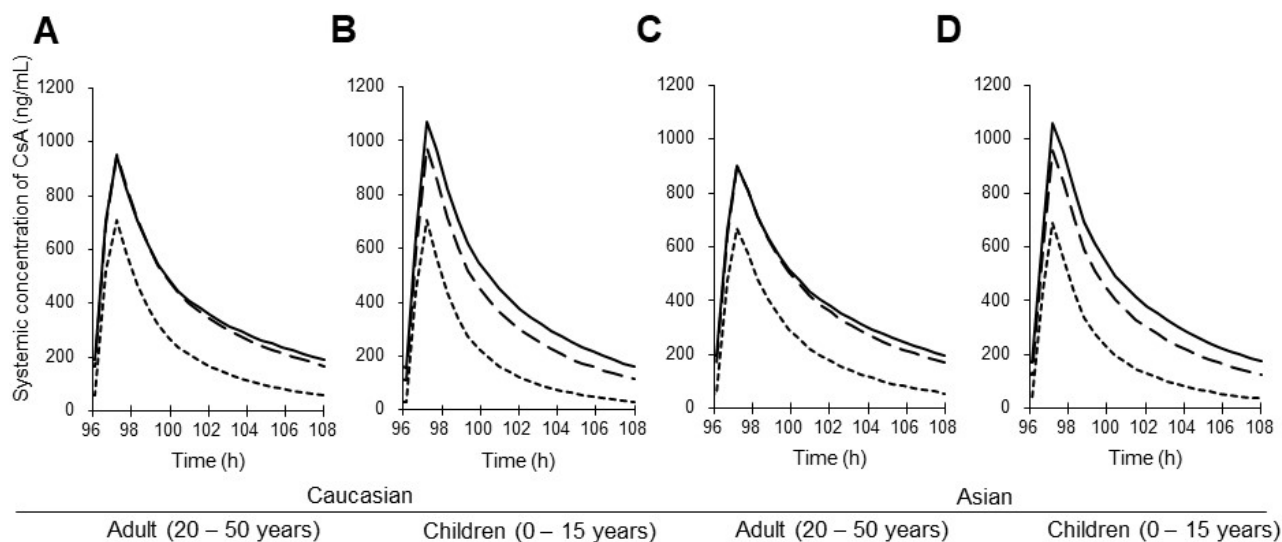


Figure 3. Comparison of PBPK model simulated steady-state plasma concentration-time profiles of cyclosporine A in (A) Caucasian adults and (B) children and (C) Asian adults and (D) children. Dotted lines indicate a healthy population. Dashed lines indicate a population with moderate renal impairment. Solid lines indicate a population with severe renal impairment.

degrees of PK parameter change between the healthy and renally impaired groups were higher in children than those of adults.

Discussion

The primary objective of this study was to identify PK of CsA in Asian children with renal impairment by means of a PBPK model. There was no corresponding built-in population in Simcyp Simulator to perform simulations. Thus, a PBPK model for Asian children with impaired renal function was developed. Final models were developed with the following processes: initial model development, verification and optimization. The lag time before absorption was modified in the optimization process. When clinical data obtained from the published literature were plotted with the simulated results, it was well-fitted within the simulation results of healthy and moderate renal impairment populations.

In addition, the coefficients of variation (CV) of observed and simulated $C_{\max,ss}$ and $AUC_{\tau,ss}$ were compared to assess the predictive performance of variability. The CV of $C_{\max,ss}$ for the PBPK model was 52–53% which were slightly higher than the CV obtained from observed data (45%). However, the CV for $AUC_{\tau,ss}$ was 92–93% which was highly overestimated when compared to the observed data (39%). The overestimation of variability in exposure could be a limitation of this PBPK model.

Furthermore, simulations with the PBPK models were conducted to assess the net influence of ethnicity and age with the disease. Slightly lower $C_{\max,ss}$ and higher $AUC_{\tau,ss}$ in Asian children compared to Caucasian children were observed. Polymorphisms of the CYP3A isozyme are known to be related to the PK of the drug. Little inter-ethnic variability of the CYP3A polymorphism between Asians and Caucasians may explain the

simulation results.[24] The age effect on the PK of CsA differed between healthy and renal impairment populations. In populations with severe renal impairment, the $C_{\max,ss}$ and $AUC_{\tau,ss}$ were both higher for children than adults, whereas healthy and moderate renally impaired children demonstrated lower $AUC_{\tau,ss}$ than healthy adults. It could be interpreted that the effect of severe renal impairment on CsA exposure is higher in children. Further clinical investigation is required to prove this hypothesis.

To our knowledge, there are limited publications or guidance on designing PBPK simulations, including the number of subjects and trials with PBPK simulations. In the validation procedure, we used the same number of subjects with observed data. For simulations examining the contribution of ethnicity, age, and disease to PK variability, the number of subjects and trials was determined considering the real-world clinical trial design. A total of 100 virtual subjects were created for each simulation as follows: 10 trials with 10 subjects per trial.

There were several limitations in this study. First, the 'Asian' population in this study needs to be more specific because Asian covers a broad spectrum of populations. However, there has been no available data for pediatric PBPK population model except Japanese. We defined our model as Asian because it was developed using two ethnic groups Japanese and Korean. Therefore, our population model could be more appropriate to the East Asian population. Second, the observed data used for validation did not contain individual GFR values. Visual inspection was consistent with that observed data were obtained from children with mean GFRs higher than 90 mL/min/1.73 m². Clinical data with individual GFR values may enable more quantitative verification of the PBPK model. In addition, the

relevance of physiological parameters modified for the newly developed PBPK model needs to be qualified by previous studies. For instance, albumin and total plasma protein levels increase from birth to 3 years. It causes alteration of unbound concentration of CsA (plasma binding >95%) and may affect the drug effect. [25,26] The change of plasma drug binding during maturation reported in recent literature was not reflected in the PBPK models in this study. For a more convincing model, however, considering quantitative physiological information is essential. Third, all of the simulation concentrations were plasma concentrations that are default in the simulator. However, the observed clinical data [20] were whole blood concentrations in the verification process. Considering that blood concentrations of cyclosporine are about 50–60% higher than their plasma values and the GFR of the children population was within normal range, the verification confirmed the healthy population model. Attention should be paid in the interpretation of Figure 1. Lastly, characterizing physiological differences by the ratio of parameters may not reflect the whole precise mechanism of disease progression. Moreover, the Simcyp Simulator uses the same transporter related parameters in both healthy and renal impairment population, which might be unrealistic. Explaining the nonlinear relationship of physiologic conditions only by ratio could be insufficient, especially for the extremes of the population. However, there is no qualified method of parameter adjustment for a specific population in PBPK modeling so far. In this state, multiplying the ratio can be the most straightforward and not much biased approach we can derive. This work has a particular significance in trying this parameter adjustment strategy for the first time and comparing the simulation results with clinical data.

Furthermore, the typical PK profiles obtained from the simulation could be utilized as representative of the specific population. Accordingly, simulation results could support designing early clinical trials, such as a selection of dose and subject number. The PK profile for populations lacking in the built-in library of the Simcyp Simulator could be predicted by applying a parameter adjustment strategy used in this study. This strategy enabled identification of the net effect of several coexisting physiologic conditions by integrating into a single model. Ethical and practical issues, which are considerable in conducting clinical trials, are emphasized in particular patient populations, such as pediatric patients. Modeling and simulation enable the assessment of the mean parameter estimates of the specific populations; thus, it could be highly utilized for populations with difficulties in conducting real-world clinical trials.

In summary, the developed PBPK model of Asian children with renal impairment adequately characterized the PK profiles of CsA. Simulations with populations categorized by ethnicity, age, and renal function enabled the assessment of PK differences between each population. This model may be a useful tool to predict the PK of CsA and support dose adjustment or other relevant decision-making in clinical settings.

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Conflict of interest

- Authors: Nothing to declare
- Reviewers: Nothing to declare
- Editors: Nothing to declare

References

1. Henriques Ldos S, Matos Fde M, Vaisbich MH. Pharmacokinetics of cyclosporine - a microemulsion in children with idiopathic nephrotic syndrome. *Clinics (Sao Paulo)* 2012;67:1197-1202. doi: 10.6061/clinics/2012(10)12.
2. Hricik DE, O'Toole MA, Schlak JA, Herson J. Steroid-free immunosuppression in cyclosporine-treated renal transplant recipients: a meta-analysis. *J Am Soc Nephrol* 1993;4:1300-1305. doi: 10.1034/j.1600-6143.2002.020105.x.
3. Kidney Disease: Improving Global Outcomes (KDIGO) Glomerulonephritis Work Group. KDIGO Clinical Practice Guideline for Glomerulonephritis. *Kidney Inter* 2012; 2:S139-S274. doi: 10.1038/kisup.2012.10.
4. Clardy CW, Schroeder TJ, Myre SA, Wadhwa NK, Pesce AJ, First MR, et al. Clinical variability of cyclosporine pharmacokinetics in adult and pediatric patients after renal, cardiac, hepatic, and bone-marrow transplants. *Clin Chem* 1988;34:2012-2015.
5. del Mar Fernández De Gatta M, Santos-Buelga D, Domínguez-Gil A, García MJ. Immunosuppressive therapy for paediatric transplant patients: pharmacokinetic considerations. *Clin Pharmacokinet* 2002;41:115-135.
6. Min DI, Ellingrod VL, Marsh S, McLeod H. CYP3A5 polymorphism and the ethnic differences in cyclosporine pharmacokinetics in healthy subjects. *Ther Drug Monit* 2004;26:524-528.
7. Lindholm A, Welsh M, Alton C, Kahan BD. Demographic factors influencing cyclosporine pharmacokinetic parameters in patients with uremia: racial differences in bioavailability. *Clin Pharmacol Ther* 1992;52:359-371.
8. Kimchi-Sarfaty C, Marple AH, Shinar S, Kimchi AM, Scavo D, Roma MI, et al. Ethnicity-related polymorphisms and haplotypes in the human ABCB1 gene. *Pharmacogenomics* 2007;8:29-39. doi: 10.2217/14622416.8.1.29.
9. Christians U, Strom T, Zhang YL, Steudel W, Schmitz V, Trump S, et al. Active drug transport of immunosuppressants: new insights for pharmacokinetics and pharmacodynamics. *Ther Drug Monit* 2006;28:39-44.
10. Lalonde L, Charpiat B, Leboucher G, Tod M. Consequences of renal failure on non-renal clearance of drugs. *Clin Pharmacokinet* 2014;53:521-532. doi: 10.1007/s40262-014-0146-1.
11. Nolin TD. Altered nonrenal drug clearance in ESRD. *Curr Opin Nephrol Hypertens* 2008;17:555-559. doi: 10.1097/MNH.0b013e3283136732.
12. Yee GC, Lennon TP, Gmur DJ, Kennedy MS, Deeg HJ. Age-dependent cyclosporine: pharmacokinetics in marrow transplant recipients. *Clin Pharmacol Ther* 1986;40:438-443.
13. Burckart G, Starzl T, Williams L, Sanghvi A, Gartner C, Venkataramanan R, et al. Cyclosporine monitoring and pharmacokinetics in pediatric liver transplant patients. *Transplant Proc* 1985;17:1172-1175.
14. Cooney GF, Habucky K, Hoppu K. Cyclosporin pharmacokinetics in pae-

- diatric transplant recipients. *Clin Pharmacokinet* 1997;32:481-495. doi: 10.2165/00003088-199732060-00004.
15. Yee GC, Lennon TP, Gmur DG, Carlin J, Schaffer RL, Kennedy MS, et al. Clinical pharmacology of cyclosporine in patients undergoing bone marrow transplantation. *Transplant Proc* 1986;18(6 Suppl 5):153-159.
 16. Sandimmune® (cyclosporine) capsules [package insert]. East Hanover (NJ): Novartis Pharmaceuticals Corporation; 2015. [cited 2017 Sep 05]. Available from: <https://www.pharma.us.novartis.com/sites/www.pharma.us.novartis.com/files/sandimmune.pdf>
 17. Jin M, Seto W, Taylor T, Saunders EF, Doyle J, Dupuis LL. Determination of initial i.v. CYA dosage to achieve target AUC values in pediatric hematopoietic stem cell transplant patients. *Bone Marrow Transplant* 2008;42:455-459. doi: 10.1038/bmt.2008.189.
 18. Krauss M, Tappe K, Schuppert A, Kuepfer L, Goerlitz L. Bayesian Population Physiologically-Based Pharmacokinetic (PBPK) Approach for a Physiologically Realistic Characterization of Interindividual Variability in Clinically Relevant Populations. *PLoS One* 2015;10:e0139423. doi: 10.1371/journal.pone.0139423.
 19. Zhao P, Rowland M, Huang SM. Best practice in the use of physiologically based pharmacokinetic modeling and simulation to address clinical pharmacology regulatory questions. *Clin Pharmacol Ther* 2012;92:17-20. doi: 10.1038/clpt.2012.68.
 20. Chun WS. Pharmacokinetics of Cyclosporine A and Its Therapeutic Effect in Children with Renal Diseases. *Korean J Pediatr* 2004;47:193-203.
 21. Maharaj AR, Barrett JS, Edginton AN. A workflow example of PBPK modeling to support pediatric research and development: case study with lorazepam. *AAPS J* 2013;15:455-464. doi: 10.1208/s12248-013-9451-0.
 22. Edginton AN, Schmitt W, Willmann S. Development and evaluation of a generic physiologically based pharmacokinetic model for children. *Clin Pharmacokinet* 2006;45:1013-1034. doi: 10.2165/00003088-200645100-00005.
 23. Cremers SC, Scholten EM, Schoemaker RC, Lentjes EG, Vermeij P, Paul LC, et al. A compartmental pharmacokinetic model of cyclosporin and its predictive performance after Bayesian estimation in kidney and simultaneous pancreas-kidney transplant recipients. *Nephrol Dial Transplant* 2003;18:1201-1208.
 24. Kurose K, Sugiyama E, Saito Y. Population differences in major functional polymorphisms of pharmacokinetics/pharmacodynamics-related genes in Eastern Asians and Europeans: implications in the clinical trials for novel drug development. *Drug Metab Pharmacokinet* 2012;27:9-54. doi: 10.2133/dmpk.DMPK-11-RV-111.
 25. Schmidt S, Gonzalez D, Derendorf H. Significance of protein binding in pharmacokinetics and pharmacodynamics. *J Pharm Sci* 2010;99:1107-1122. doi: 10.1002/jps.21916.
 26. Sethi PK, White CA, Cummings BS, Hines RN, Muralidhara S, Bruckner JV. Ontogeny of plasma proteins, albumin and binding of diazepam, cyclosporine, and deltamethrin. *Pediatr Res* 2016;79:409-415. doi: 10.1038/pr.2015.237.
 27. Rodgers T, Rowland M. Mechanistic approaches to volume of distribution predictions: understanding the processes. *Pharm Res* 2007;24:918-933. doi: 10.1007/s11095-006-9210-3.